BIOLOGICAL ACTIVITY ASSESSMENT OF 25-HYDROXYVITAMIN D₃-26,23-LACTONE IN THE RAT

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Received 20 January 1982

1. Introduction

A new metabolite of vitamin D₃ has been isolated from the plasma of chickens, rats and pigs given large doses of vitamin D₃ and identified as 25-hydroxyvitamin D_3 -26,23-lactone (25-OH- D_3 -26,23-lactone) [1-3]. Four possible diastereoisomers of 25-OH-D₃-26,23-lactone have been synthesized [4-9]; they have been directly compared to the natural 25-OH-D₃-26, 23-lactone by high pressure liquid chromatography. The stereochemical configurations of the natural 25-OH-D₃-26,23-lactone at C-23 and C-25 positions were determined to be 23(S) and 25(R), respectively [9,10]. 23(S)25-Dihydroxyvitamin D_3 [23(S)25- $(OH)_2D_3$] is a far better substrate for production of 25-OH-D₃-26,23-lactone than is 25,26-dihydroxyvitamin D_3 (25,26-(OH)₂ D_3) [11]. 23(S)25(R)26-trihydroxyvitamin D_3 [23(S)25(R)26-(OH)₃ D_3] is a better substrate than $23(S)25-(OH)_2D_3$ in the biosynthesis of the 25-OH-D₃-26,23-lactone [12]. Therefore, the 25-OH-D₃-26,23-lactone may be biosynthesized from 25-OH-D₃ by way of 23(S)25-(OH)₂D₃ to 23(S)25- $(R)26-(OH)_3D_3$.

The biological activity and physiological function(s) of the 25-OH—D₃-26,23-lactone are still unknown. Here, we report that in the rat that 25-OH-D₃-26,23-lactone slightly stimulates intestinal calcium transport and at the same time mediates a fall in the concentration of serum calcium.

Abbreviations: 25-OH-D₃, 25-hydroxyvitamin D₃; 25-OH-D₃-26,23-lactone, 25-hydroxyvitamin D₂-26,23 lactone; 23,25-(OH)₂D₃, 23,25-dihydroxyvitamin D₃; 25,26-(OH)₂D₃, 25,26-dihydroxyvitamin D₃; 1α ,25-(OH)₂D₃, 1α ,25-dihydroxyvitamin D₃; 23,25,26-(OH)₃D₃, 23,25,26-trihydroxyvitamin D₃; 1α , 25-(OH)₂D₃-26,23-lactone, 1α ,25-dihydroxyvitamin D₃-26,23-lactone

2. Materials and methods

2.1. Compounds

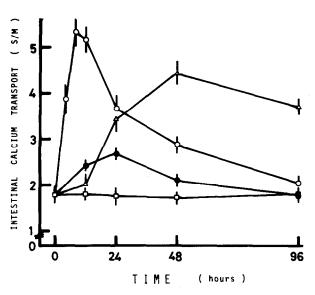
We synthesized 25-OH-D₃ and 1α ,25-(OH)₂D₃ as in [13]. 25-OH-D₃-26,23-Lactone was isolated and purified from the serum of rats given large doses of vitamin D₃ as in [10].

2.2. Assay for intestinal calcium transport and bone calcium mobilization

Male weanling rats of the Wistar strain were fed a vitamin D-deficient low calcium diet (Ca, 0.0036%; P, 0.3%; Teklab Test Diet, Madison WI) for 6 weeks. At the end of week 6, 5 rats each (\sim 100 g body wt) received an intravenous injection of 500 ng 25-OH-D₃-26,23-lactone, 250 ng 1α ,25-(OH)₂D₃ or 500 ng 25-OH-D₃ in 0.2 ml 0.2% Triton X-100 solution. The rats were sacrificed at the indicated times after administration, and the intestinal calcium transport and serum calcium concentration were measured. The intestinal calcium transport assay using everted duodenal sacs was done as in [14]. Serum calcium concentration was determined by the OCPC (o-cresol-phthalein complexone) method [15].

3. Results

The biological activity of the isolated 25-OH-D₃-26,23-lactone was assayed in vitamin D-deficient rats fed a low calcium diet. Initially, a single dose of 500 ng 25-OH-D₃-26,23-lactone was used to test its intestinal calcium transport activity in comparison to that of 1α ,25-(OH)₂D₃ or 25-OH-D₃ (fig.1). The 25-OH-D₃-26,23-lactone slightly stimulated intestinal calcium transport after 12 h and it reached a maxi-



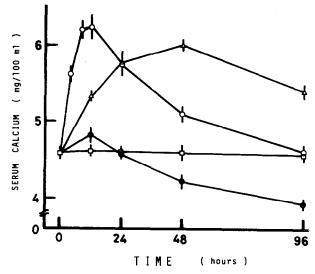


Fig. 1. Time-course response of intestinal calcium-transport system. Rats on a low calcium diet received a single intravenous dose of either 500 ng 25-OH-D₃-26,23-lactone (\bullet) or 250 ng 1α ,25-(OH)₂D₃ (\circ) or 500 ng 25-OH-D₃ (\triangle) or vehicle (\square) in 0.2 ml 0.2% Triton X-100 solution. At the indicated times, animals were decapitated and their duodena were used for the determination of intestinal calcium transport. The rate of intestinal calcium transport is represented by the ratio of $^{45}\text{Ca}^{2+}$ in the serosal medium to $^{45}\text{Ca}^{2+}$ in the mucosal medium. Each point is the mean \pm SEM of determinations on 5 rats.

Fig. 2. Time-course response of bone calcium mobilization induced in rats by 500 ng 25-OH-D₃-26,23-lactone (\bullet) or 250 ng 1α ,25-(OH)₂D₃ (\circ) or 500 ng 25-OH-D₃ (\triangle) or vehicle (\circ). Rats on a low calcium diet received a single intravenous dose of compound in 0.2 ml 0.2% Triton X-100 solution. At the indicated times, animals were decapitated, blood was collected, and calcium was measured in the serum by the OCPC method [15]. Data are expressed as mg Ca²⁺/100 ml serum and are the mean \pm SEM of 5 determinations.

mum 24 h after administration. After 48 h, the responses did not differ significantly from the control. On the contrary, $1\alpha,25$ -(OH)₂D₃ and 25-OH-D₃ demonstrated a greater stimulation of intestinal calcium transport than did 25-OH-D₃-26,23-lactone.

The time course of calcium mobilization from bone, as measured by an elevation in serum ${\rm Ca^{2+}}$ levels, produced by 500 ng 25-OH-D₃-26,23-lactone was compared to that produced by 250 ng 1α ,25-(OH)₂D₃ or 500 ng 25-OH-D₃ (fig.2). The 25-OH-D₃-26,23-lactone caused only a slight elevation in serum calcium over the first 24 h of assay. This was followed over the 48–96 h interval by a significant reduction in the serum calcium levels. However, 1α ,25-(OH)₂D₃ and 25-OH-D₃ generate a significant rise in serum ${\rm Ca^{2+}}$ levels, in accord with [18,19], demonstrating potent actions of these *seco*-steroids on bone calcium mobilization.

Table 1 shows the dose—response relationship between 25-OH-D₃-26,23-lactone and intestinal calcium transport and bone calcium mobilization. The intestinal calcium transport of 1α ,25-(OH)₂D₃ reached

a maximum within 8 h after dosing, while that of 25-OH-D₃-26,23-lactone only reached a maximum 24 h after administration (fig.1). The more 25-OH-D₃-26,23-lactone is administered, the more intestinal calcium transport is stimulated. The potency of 25-OH-D₃-26,23-lactone in intestinal calcium transport was estimated to be \sim 120-times less than that of 1α ,25-(OH)₂D₃. Graded doses of 25-OH-D₃-26,23-lactone had no influence on bone calcium mobilization 24 h after dosing.

Table 2 illustrates the effect of prior dosing with 25-OH-D₃-26,23-lactone on the biological response to a subsequently administered dose of 1,25-(OH)₂D₃. In agreement with the results in fig.1,2, 95 h after the administration of the 25-OH-D₃-26,23-lactone alone, there was apparent a significant inhibition of bone calcium mobilization. Intriguingly, the prior administration of 25-OH-D₃-26,23-lactone completely blocked the characteristic bone calcium mobilization response of 1,25-(OH)₂-D₃, while it had no inhibiting effect on the intestinal calcium transport of 1α ,25-(OH)₂D₃.

Table 1

Dose—response relationship between 25-OH-D₃-26,23-lactone and intestinal calcium transport and bone calcium mobilization in vitamin D-deficient rats on a low calcium diet

| Compound | Dose (ng) | Time (h) | ⁴⁵ Ca ²⁺ serosal/ ⁴⁵ Ca ²⁺ mucosal | Serum Ca ²⁺ (mg/100 ml) |
|--|--------------|-------------|---|---------------------------------------|
| Vehicle | | 8 | 1.74 ± 0.16 | 4.74 ± 0.17 |
| 1α,25-(OH) ₂ D ₃ | 10 | 8 | 2.48 ± 0.15^{a} | 4.95 ± 0.18^{a} |
| | 50 | 8 | $4.20 \pm 0.22^{\circ}$ | $5.60 \pm 0.18^{\circ}$ |
| | 100 | 8 | $5.06 \pm 0.31^{\circ}$ | $5.92 \pm 0.20^{\circ}$ |
| | 250 | 8 | $5.33 \pm 0.28^{\circ}$ | $6.09 \pm 0.22^{\circ}$ |
| 25-OH-D ₃ -26,23-lactone | 100 | 24 | 2.14 ± 0.13^{a} | 4.80 ± 0.16 |
| | 500 | 24 | 2.65 ± 0.20^{b} | 4.70 ± 0.18 |
| | 2500 | 24 | 3.27 ± 0.24^{b} | 4.62 ± 0.21 |

Significantly different from control: a p < 0.05; b p < 0.01; c p < 0.001

After 6 weeks on the vitamin D-deficient, low calcium diet, rats were divided into groups of 3 or 5 and each rat received a single intravenous dose of compound in 0.2 ml 0.2% Triton X-100 solution. Control rats received only vehicle. After 8 or 24 h, the animals were decapitated and intestinal calcium transport and serum calcium concentration were measured as in section 2. Data are expressed as mean \pm SEM

4. Discussion

We report the first detailed assessment of the biological activity of the characterized vitamin D_3 metabolite, 25-OH- D_3 -26,23-lactone. We have employed biosynthetically produced 25-OH- D_3 -26,23-lactone, independently shown to have the stereochemical configuration of 23(S) and 25(R) [9,10]. These studies in the rat indicate that the naturally occurring 25-OH- D_3 -26,23-lactone has <1% (table 1) of the activity of

1,25-(OH)₂-D₃ in stimulating intestinal calcium transport and no capability to stimulate bone calcium mobilization (fig.1,2). The incubation of rachitic chicken kidney homogenates with 25-OH-D₃-26,23-lactone is known to produce substantial amounts of 1α -25,-dihydroxyvitamin-D₃-26,23-lactone [1,25-(OH)₂-D₃-26,23-lactone] [10,17]. The biological activity of the 25-OH-D₃-26,23-lactone may result from enzymatic conversion of this seco-steroid into the 1,25-(OH)₂-D₃-26,23-lactone.

Table 2

Effect of 25-OH-D₃-26,23-lactone on intestinal calcium transport and serum calcium concentration of vitamin D-deficient rats

| Compound given | | Intestinal Ca ²⁺ transport | Serum Ca ²⁺ |
|---|------------------------|---|------------------------|
| Dose 1 | Dose 2 | 45Ca ²⁺ serosal/45Ca ²⁺ mucosal | (mg/100 ml) |
| Vehicle 25-OH-D ₂ -26,23- | Vehicle | 1.71 ± 0.13 | 4.84 ± 0.18 |
| lactone | Vehicle | 1.80 ± 0.09 | 4.14 ± 0.19^{a} |
| Vehicle 25-OH-D ₃ -26,23- | $1\alpha,25-(OH)_2D_3$ | 3.96 ± 0.18^{a} | 5.82 ± 0.16^{a} |
| lactone | $1\alpha,25-(OH)_2D_3$ | 3.91 ± 0.10^{a} | 4.88 ± 0.20 |

Significantly different from control: a p < 0.001

Rats fed a low calcium, vitamin D-deficient diet for 6 weeks into 4 groups of 5 rats. They received the first dose of 500 ng 25-OH-D₃-26,23-lactone dissolved in 0.2 ml 0.2% Triton X-100 solution or vehicle only intravenously. After 80 h they received a second dose of 250 ng 1α ,25-(OH)₂D₃ dissolved in 0.2 ml 0.2% Trition X-100 solution or vehicle only by the same route; 15 h after the second dose, the animals were killed and intestinal calcium transport activity and serum calcium concentration were measured as in section 2. The data are expressed as the mean \pm SEM

The striking ability of the 25-OH-D₃-26,23-lactone to apparently inhibit bone calcium mobilization (fig.2; table 2) are novel and intriguing. Regulation of serum calcium levels in animals is known to be under a multifactorial hormonal control involving parathyroid hormone [PTH], calcitonin and 1,25-(OH)₂-D₃ [16]. Thus the observed reduction in serum calcium levels could conceivably result from an inhibition by 25-OH-D₃-26,23-lactone of PTH-mediated bone resorption or alternatively a stimulation of renal tubular calcium excretion or an inhibition of renal tubular calcium absorption. Studies are underway to evaluate these several possibilities.

Also as shown in table 2, the 25-OH-D₃-26,23-lactone has a possible significant anti-vitamin action with respect to blocking the actions of 1,25-(OH)2-D3 on bone calcium mobilization. No vitamin D analogs or metabolites had been identified which have an antivitamin activity for 1,25-(OH)₂D₃; several compounds, however, have been reported to block the biological response of vitamin D [20,21]. However, it is possible that this apparent inhibition of 1,25-(OH)₂D₃ action is simply a separate action of the 25-OH-D₃-26,23-lactone on perturbing the renal tubular handling of calcium (see above) so that the characteristic 1,25-(OH)₂-D₃-mediated elevation in serum calcium is not detectable by this experimental protocol. Future experiments will be required to evaluate how combinations of 1,25-(OH)₂-D₃ and 25-OH-D₃-26,23-lactone may regulate serum calcium levels. Only then will it be possible to gain insight into the physiological function of the 25-OH-D₃-26,23-lactone.

References

 Wichmann, J. K., DeLuca, H. F., Schnoes, H. K., Horst, R. L., Shepard, R. M. and Jorgensen, N. A. (1979) Biochemistry 18, 4775-4780.

- [2] Horst, R. L. (1979) Biochem. Res. Commun. 89, 286-293.
- [3] Horst, R. L. and Littledike, T. (1980) Biochem. Biophys. Res. Commun. 93, 149-154.
- [4] Wichmann, J. K., Paaren, H. E., Fivizzani, M. A., Schnoes, H. K. and DeLuca, H. F. (1980) Tetrahedron Lett. 21, 4667-4670.
- [5] Ikekawa, N., Hirano, Y., Ishiguro, M., Oshida, J., Eguchi, T. and Miyasaka, S. (1980) Chem. Pharm. Bull. 28, 2852-2854.
- [6] Morris, D. S., Williams, D. H. and Norris, A. F. (1981)J. Chem. Soc. Commun. 424–425.
- [7] Morris, D. S., Williams, D. H. and Norris, A. F. (1981)J. Org. Chem. 46, 3422-3428.
- [8] Yamada, S., Nakayama, K. and Takayama, H. (1981) Tetrahedron Lett. 22, 2591-2594.
- [9] Yamada, S., Nakayama, K. and Takayama, H. (1981) Chem. Pharm. Bull. 29, 2393-2396.
- [10] Ishizuka, S., Yamaguchi, H., Yamada, S., Nakayama, K. and Takayama, H. (1981) FEBS Lett. 134, 207-211.
- [11] Tanaka, Y., DeLuca, H. F., Schnoes, H. K., Ikekawa, N. and Eguchi, T (1981) Proc. Natl. Acad. Sci. USA 78, 4805-4808.
- [12] Ishizuka, S., Ishimoto, S. and Norman, A. W. (1982) 137, xxx-xxx.
- [13] Ishizuka, S., Bannai, K., Naruchi, T. and Hashimoto, Y. (1981) Steroids 37, 33-43.
- [14] Martin, D. L. and DeLuca, H. F. (1969) Am. J. Physiol. 216, 1351-1359.
- [15] Connerty, H. V. and Briggs, A. R. (1966) Am. J. Clin. Pathol. 45, 290-296.
- [16] Norman, A. W. (1979) in: Vitamin D: the Calcium Homeostatic Steroid Hormone, pp. 1-490, Academic Press, New York.
- [17] Tanaka, Y., Wichmann, J. K., Paaren, H. E., Schnoes, H. K. and DeLuca, H. F. (1980) Proc. Natl. Acad. Sci. USA 77, 6411-6414.
- [18] Wong, R. G., Myrtle, J. F., Tsai, H. C. and Norman, A. W. (1972) J. Biol. Chem. 247, 5728-5735.
- [19] Wong, R. G. and Norman, A. W. (1972) J. Nutrit. 102, 1709-1718.
- [20] Norman, A. W., Johnson, R. L. and Okamura, W. H. (1979) J. Biol. Chem. 254, 11450-11456.
- [21] Gerdes, J. M., Okamura, W. H. and Norman, A. W. (1981) Arch. Biochem. Biophys. 210, 238-245.